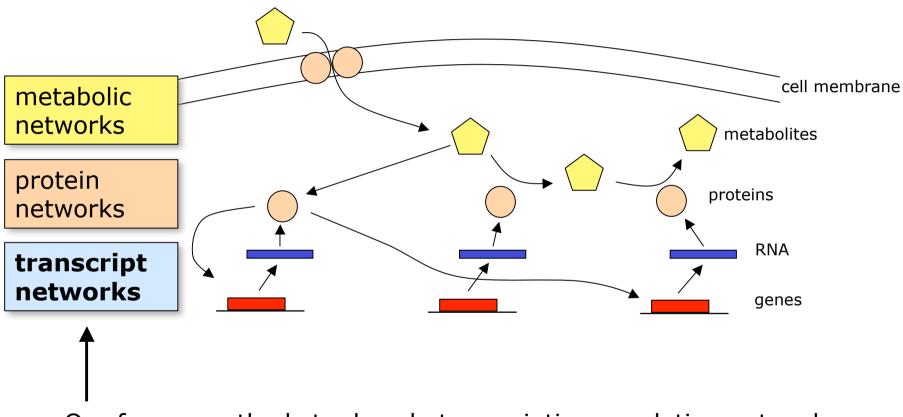
Diego di Bernardo Telethon Institute of Genetics and Medicine

19 Dicembre 2007, Napoli



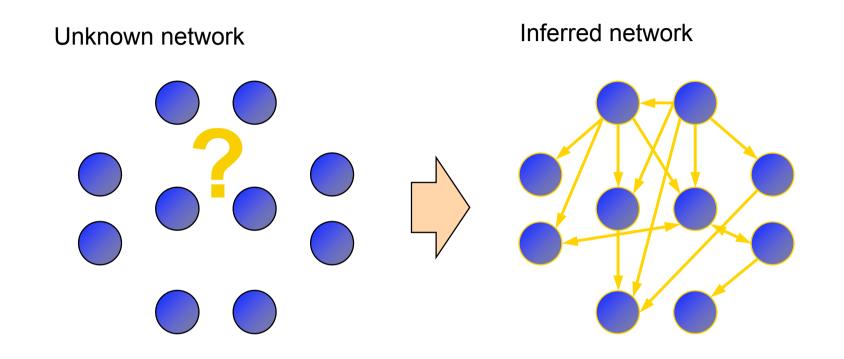
#### Gene Networks



Our focus: methods to decode transcription regulation networks

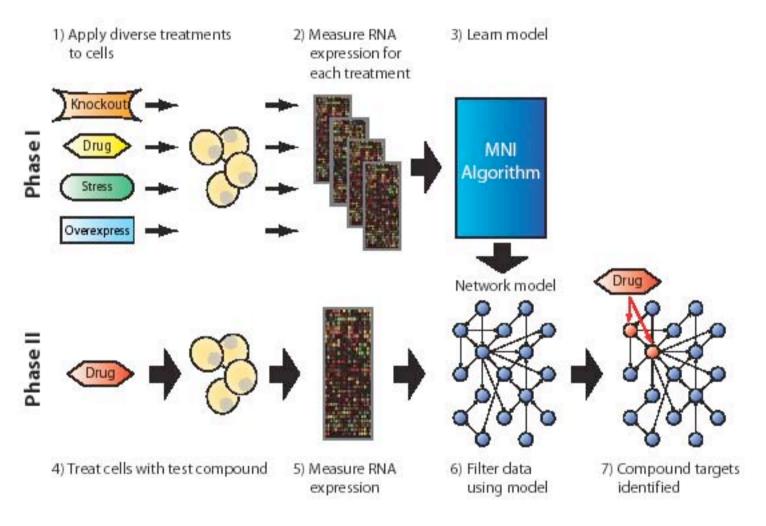


Reverse engineering (or system id) gene networks:





## Aim of reverse engineering: gene function and drug MOA

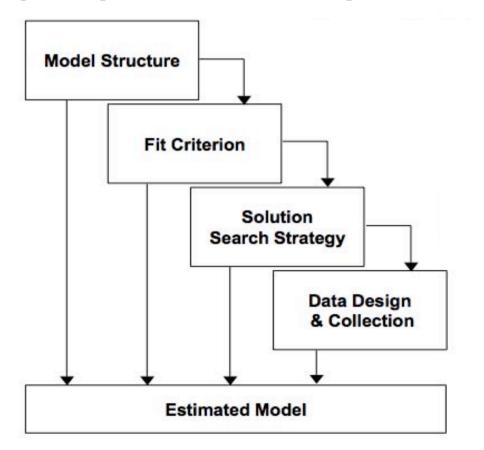


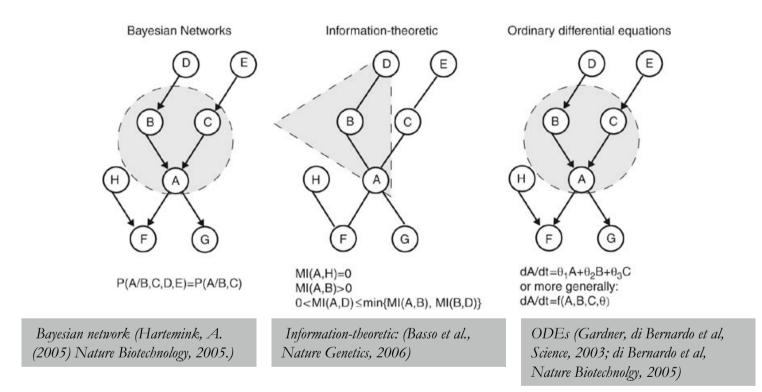


D di Bernardo et al, Nature Biotechnolgy, 2005; TS Gardner, D di Bernardo et al, Science, 2003;

#### **Reverse-engineering strategy:**

- Choose a model
- Choose a fit criterion (cost function) to measure the fit of the model to the data
- Define a strategy to find the parameters that best fit the data (i.e. that minimise cost function)
- Perform appropriate experiments to collect the experimental data:

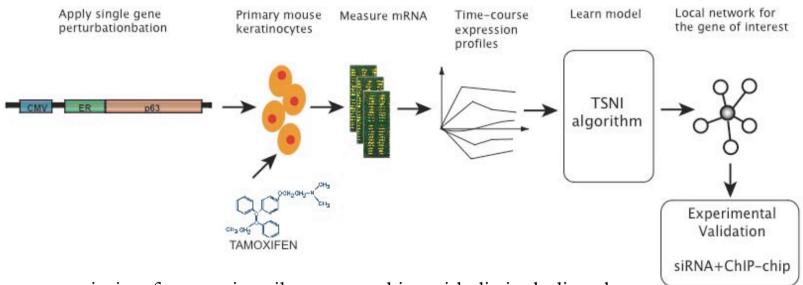




Mukesh Bansal, Vincenzo Belcastro, Alberto Ambesi, Diego di Bernardo. How to infer gene networks from expression profiles. Molecular Systems Biology, 2007.



# Understanding gene function in a genetic disease:



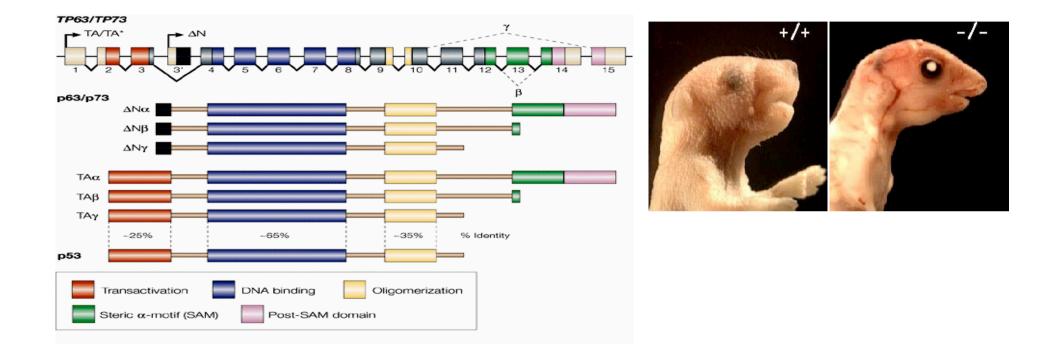
P63 is a transcription factor primarily expressed in epithelia including the proliferative compartments of the skin.

It plays an essential role in modulating cellular differentiation by unknown mechanisms.

Mutation in its DNA binding and SAM domains have been linked to five human malformation syndromes.



# The transcription factor p63:



•P63 is a Transcription Factor (unknown targets)

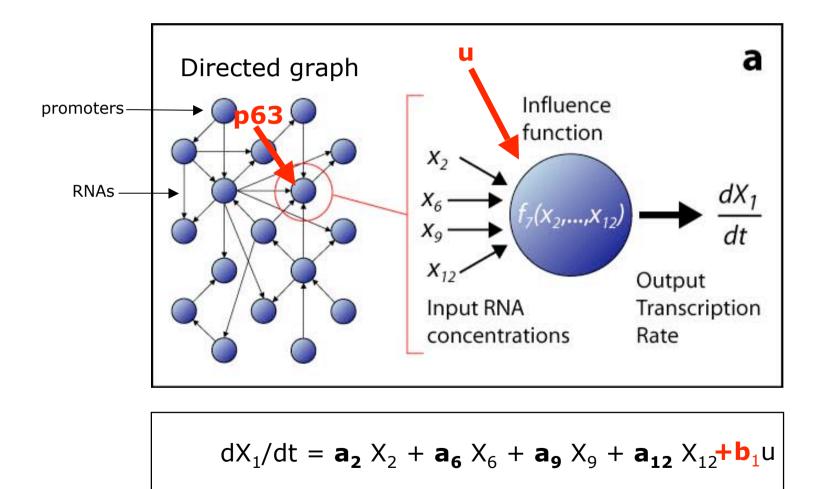
•p63 gene has three different promoter that gives rise to two different transcripts (TA and  $\Delta N$ );

•each transcript has three alternative splicing isoforms  $(\alpha, \beta, \gamma)$ 

Lack of epidermis and hair follicles
Defects in all squamous epithelia
Limb truncations
Craniofacial defects: lack of tooth primordia and eyelids
Maxilla and mandible are truncated and

secondary palate fails to close

#### Model structure: Ordinary differential equations







Model structure - Discrete model:

$$\mathbf{x}_{1}(\mathbf{t}_{k+1}) = a^{d}_{11}\mathbf{x}_{1}(\mathbf{t}_{k}) + a^{d}_{12}\mathbf{x}_{2}(\mathbf{t}_{k}) + \dots + a^{d}_{1n}\mathbf{x}_{n}(\mathbf{t}_{k}) + b^{d}_{1}\mathbf{u}(\mathbf{t}_{k})$$

$$\dots$$

$$\mathbf{x}_{n}(\mathbf{t}_{k+1}) = a^{d}_{n1}\mathbf{x}_{1}(\mathbf{t}_{k}) + a^{d}_{n2}\mathbf{x}_{2}(\mathbf{t}_{k}) + \dots + a^{d}_{nn}\mathbf{x}_{n}(\mathbf{t}_{k}) + b^{d}_{n}\mathbf{u}(\mathbf{t}_{k})$$

Or in matrix format:

$$\mathbf{x}(\mathbf{t}_{k+1}) = \mathbf{A}^{d}\mathbf{x}(\mathbf{t}_{k}) + \mathbf{b}^{d}\mathbf{u}(\mathbf{t}_{k})$$

For k=1..M

$$X(t_{k+1}) = A^{d}X(t_{k}) + b^{d}u(t_{k})$$

Where (NxM)=(NxN)(NxM)+(Nx1)(1xM)



$$\dot{X}(t_k) = AX(t_k) + BU(t_k) \quad k = 1 \dots M$$

$$X(t_{k+1}) = A_d * X(t_k) + B_d * U(t_k)$$

$$X(t_{k+1}) = \begin{bmatrix} A_d & B_d \end{bmatrix} * \begin{bmatrix} X(t_k) \\ U(t_k) \end{bmatrix}$$

NxM = Nx(N+1) (N+1)xM $X(t_{k+1}) = H * Y(t_k)$ 

$$B = (A_d + 1)^{-1} B_d$$

$$H = \begin{bmatrix} A_d & B_d \end{bmatrix}$$
$$Y(t_k) = \begin{bmatrix} X(t_k) \\ U(t_k) \end{bmatrix}$$



- Usually N>>M (more unknown than data points; i.e. 1000 genes e 20 points)
- From the previous equation we know that a unique solution will exist only if N≤M-1
- We need a trick to reduce number of unknowns by dimensional reduction:
  - Regression with Variable selection
  - Clustering
  - Regression with SVD



 $X(t_{k+1}) = H * Y(t_k)$  $H = \begin{bmatrix} A_d & B_d \end{bmatrix}$  $Y(t_k) = \begin{bmatrix} X(t_k) \\ U(t_k) \end{bmatrix}$ 

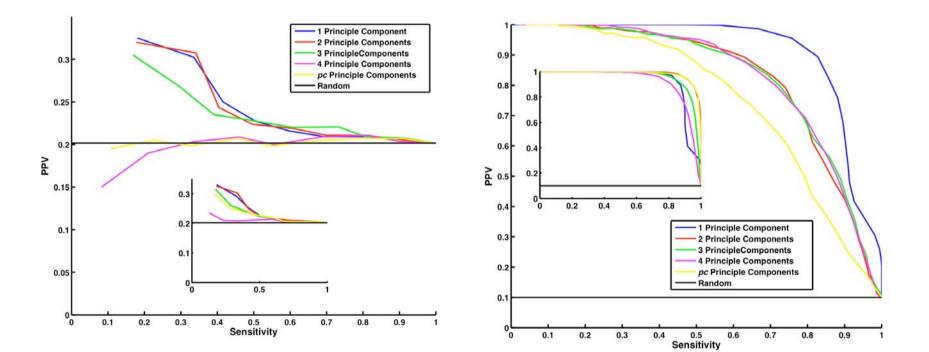
- Singular Value Decomposition:
  - Y=UDV' (N+1×M)=(N+1×M)(M×M)(M×M)

- $Y=U_rD_rV_r'$  (N+1xM)=(N+1xK)(KxK)(KxM)
- $X(t_{k+1}) = HY = (N \times N + 1)(N + 1 \times M) =$
- $X(t_{k+1}) = HU_rD_rV_r' = (HU_r)D_rV_r' = (NxK)(KxK)(KxM)$



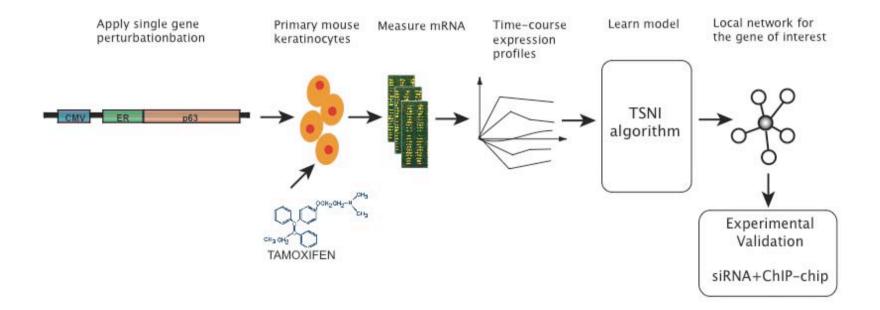
Network of 10 genes (A)

# Target prediction in 1000 gene network (B)

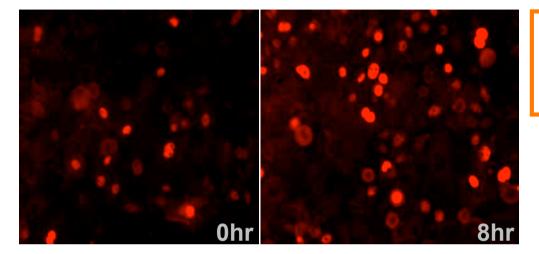




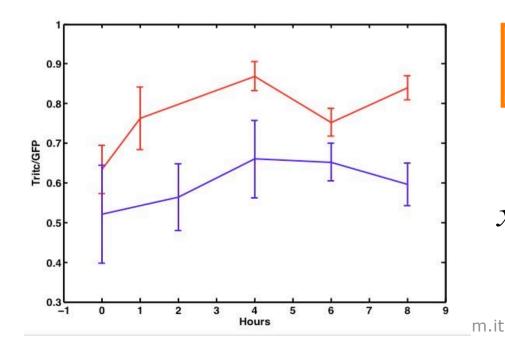
# Application to p63:







Murine keratinocytes are infected with Ginco ErDNp63 retrovirus. The cells are collected at the following time points from Tamoxifen induction: 1hr, 2hrs,4hrs, 8hrs.

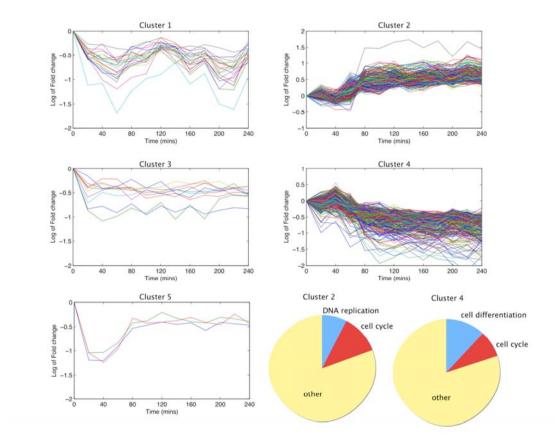


Immunofluorescence plot: red and blue lines represent two different immunofluorescence experiments.

 $\dot{x}(t) = A \cdot x(t) + Bu(t)$ 

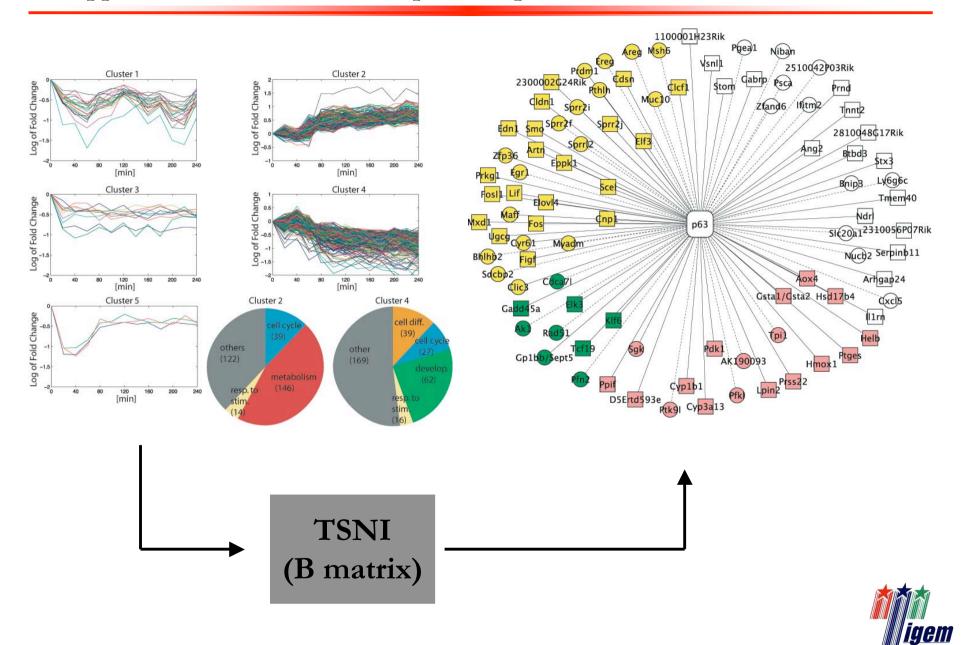


# Hierarchical Clustering of 786 expression profiles:





# Application of TSNI on 786 expression profiles:



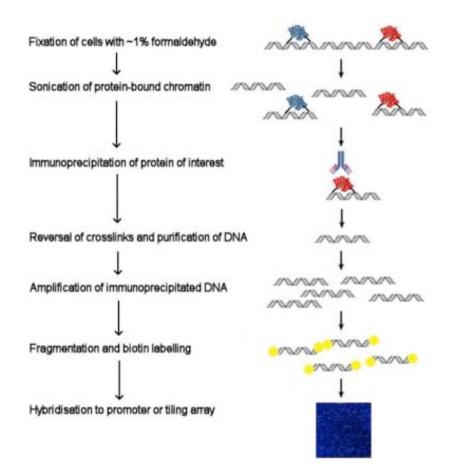
• We selected the top 100 predicted targets and as control the bottom 200 predicted genes.

•For each gene we analysed 20 Kbp upstream+gene sequence via ChIP-chip (Agilent 200k custom-array)

•We found about 300 p63-binding sites bu ChIP-chip with a p-value<0.01

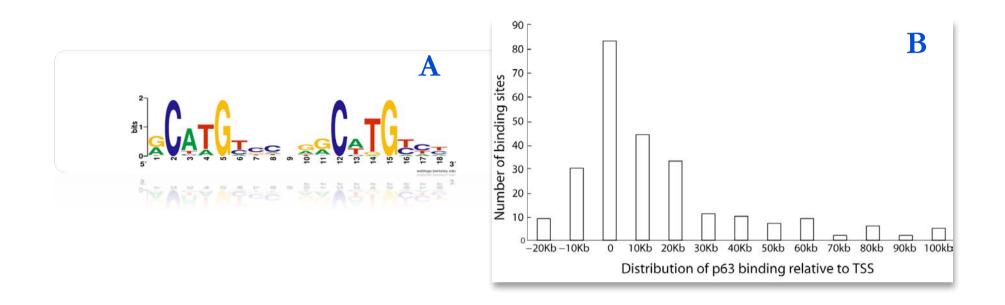


# Validation of predicted targets: (1) ChIP-chip

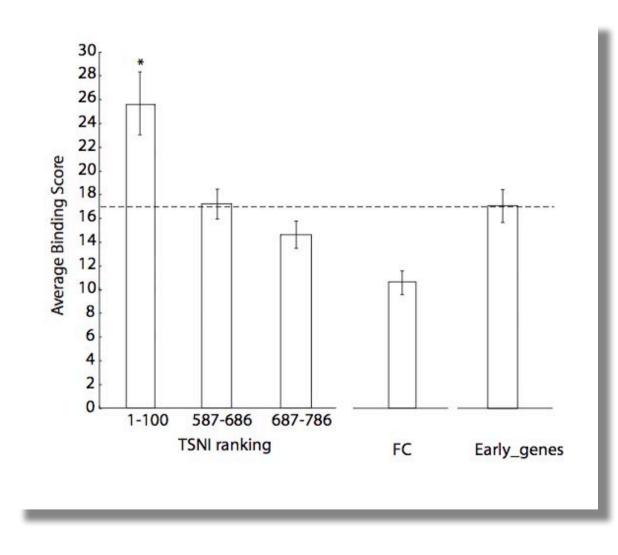




# Validation of predicted targets: (1) ChIP-chip



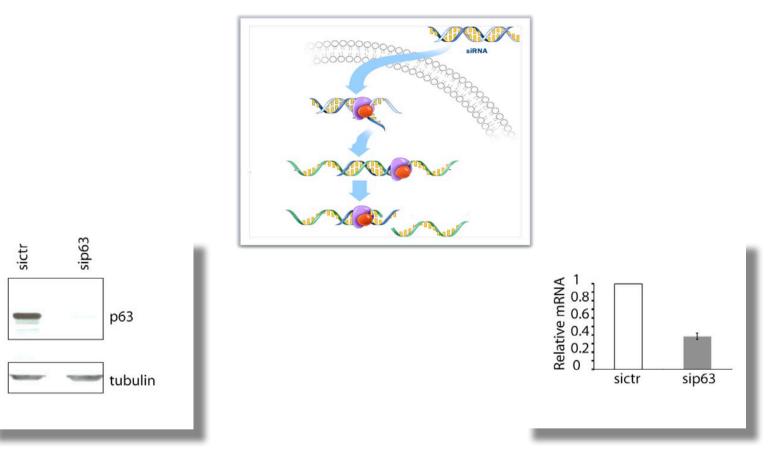






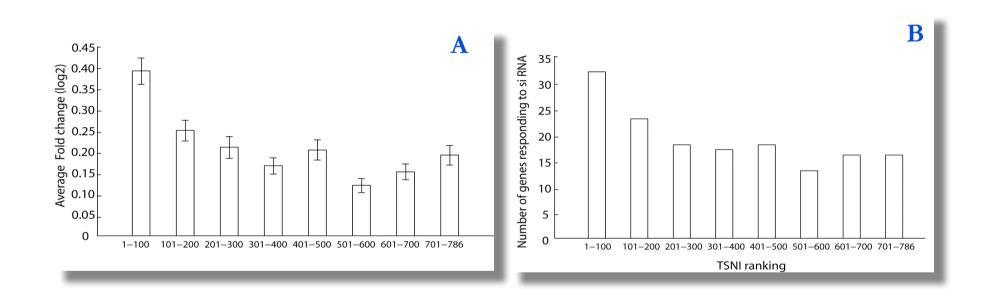
# Knocking down p63 expression via siRNA:

# RNA interference against the p63 DNA binding domain



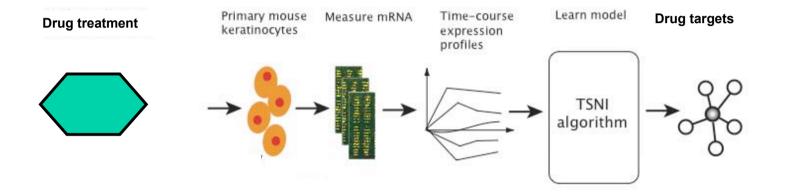


# TSNI results are confirmed by siRNA



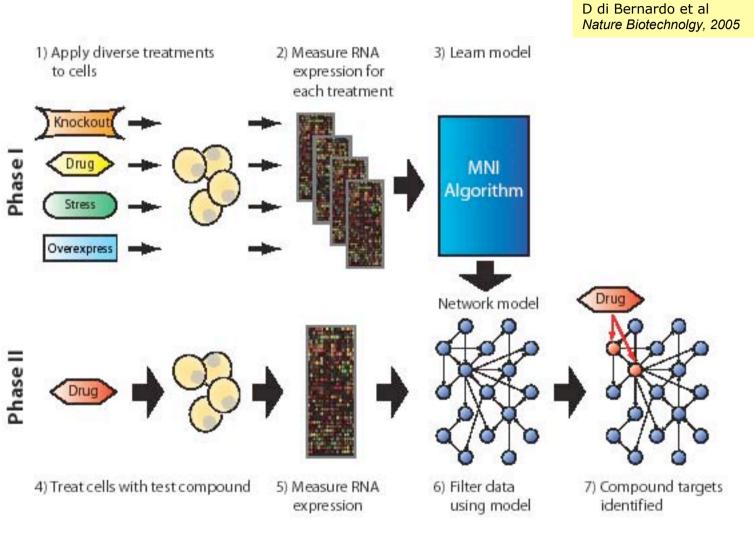


# Application of TSNI to drug discovery :





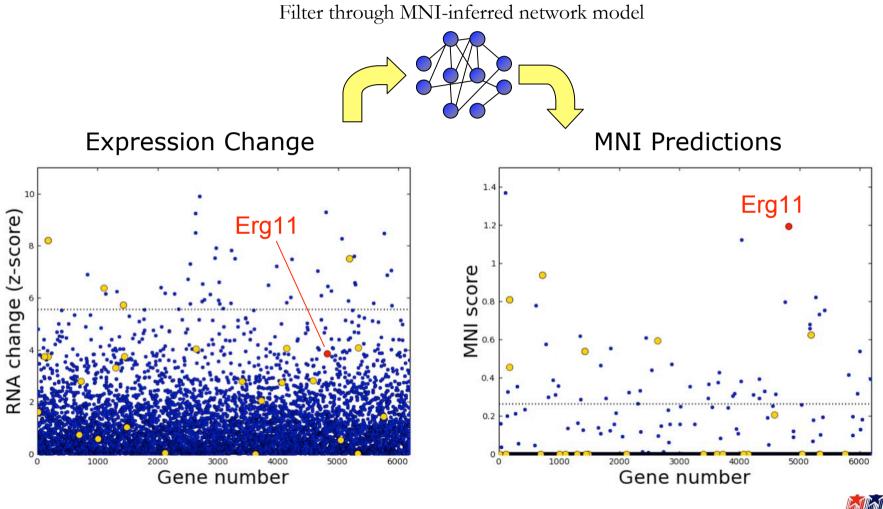
# A "steady-state" approach to drug target identification:





# MNI identifies target of itraconazole in S. cerevisiae

Itraconazole treatment: a known target is ERG11





Compound	Known pathway	Known target	Predicted pathway	Ranked target genes (rank)
ltraconazole	ergosterol biosynthesis	Erg I I	steroid metabolism	<b>ERGII,</b> ERG24, ERG1, ERG25, CYB5, ERG27, ATF2
			, in the second	2     4     6     13       16     19     23
Hydroxyurea	DNA replication	Rnr2, Rnr4	DNA replication	<b>RNR4, RNR2,</b> RNR1, RNR3
	<b>X</b>			2 6 14 23

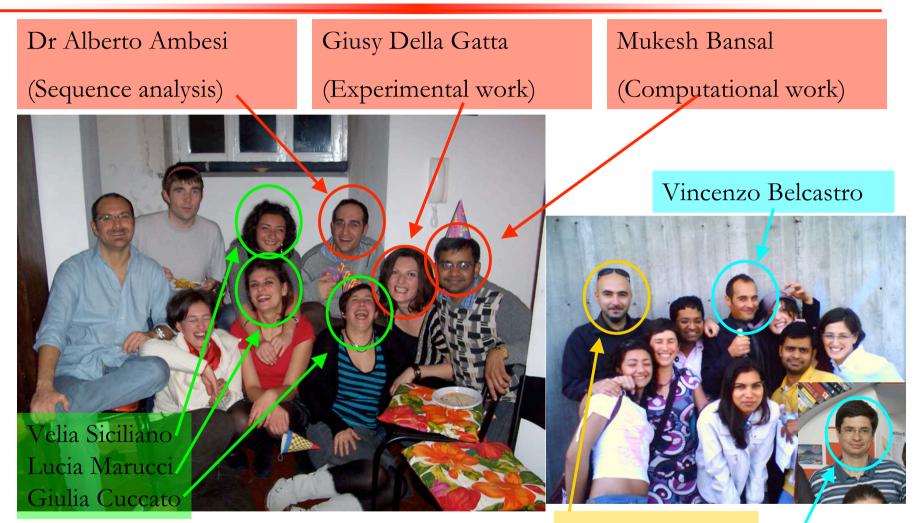
Pathway data from GO database



Drug	Known pathway	Known target	Significant GO ontology (rank, <i>P</i> -value)	Ranked pathway genes (rank)
Terbinafine	Ergosterol biosynthesis <sup>41</sup>	Ergl	Steroid metabolism (1, 10 <sup>-14</sup> )	ERG7 (4), ERG1 (5), ERG8 (11), ERG26 (13), UPC2 (17), ERG28 (18), ERG11 (20), DAP1 (33), HES1 (34) ATF2 (36), ERG5 (49)
Lovastatin	Ergosterol biosynthesis <sup>42</sup>	Hmg2, Hmg1	Lipid metabolism (1, 10 <sup>-4</sup> )	BST1 (1), ERG1 (18), YSR3 (23), <b>HMG2 (30)</b> , LCB5 (31), ERG13 (36), VRG4 (48)
Itraconazole	Ergosteroll biosynthesis <sup>43</sup>	Erg11	Steroid metabolism (1, 10 <sup>-8</sup> )	<b>ERG11 (2)</b> , ERG24 (4), ERG1 (6), ERG25 (13), CYB5 (16), ERG27 (19), ATF2 (23)
Hydroxyurea	DNA replication <sup>44</sup>	Rnr2, Rnr4	Heteroduplex formation (1, 10 <sup>-4</sup> )	RAD51 (15), RAD54 (47)
			DNA replication (2, 10 <sup>-2</sup> )	<b>RNR4 (2), RNR2 (6)</b> , RNR1 (14), RNR3 (23)
Cycloheximide	Protein biosynthesis <sup>45</sup>	Ribosome	Nuclear mRNA splicing, via spliceosome (1, 10 <sup>-4</sup> )	SYF1 (3), SMD3 (19), HSH49 (42)
			1 <del></del>	RPL26B (32), RPS29A (34)
Tunicamycin	N-linked glycosylation <sup>46</sup>	Alg7	Protein-ER targeting (1, 10 <sup>-3</sup> )	SEC62 (1), SIL1 (31), SEC59 <sup>a</sup> (43)
Nikkomycin	Cell wall chitin biosynthesis <sup>47</sup>	Chs3	Protein amino acid alkylation(1, 10 <sup>-3</sup> )	SWD2 (3), RMT2 (6)
		Drugs not in th	e original compendium data s	et
3-aminotriazole	Histidine biosynthesis <sup>48</sup>	His3	Organic acid metabolism (1, 10 <sup>-7</sup> )	FRM2 (8), BIO5 (9), YAT2 (10), ARO10 (18), ARO9 (20), CHA1 (21), BIO3 (31), ARG1 (33), ARG4 (37), HIS5 <sup>†</sup> (42), LYS1 (47), SAM2 (50)
	Oxygen and reactive oxygen species metabolism <sup>30</sup>	Ctal		
Dyclonine	Ergosterol biosynthesis <sup>1</sup>	Erg2	Sterol biosynthesis (1, 10 <sup>-18</sup> )	ERG3 (1), ERG6 (2), CYB5 (3), ERG2 (4), ERG11 (6), ERG28 (10), ERG1 (12), ERG5 (13), ERG27 (18), MVD1 (23), ERG24 (30), ERG26 (37)



# Acknowledgements (http://dibernardo.tigem.it for more info):



Francesco Iorio

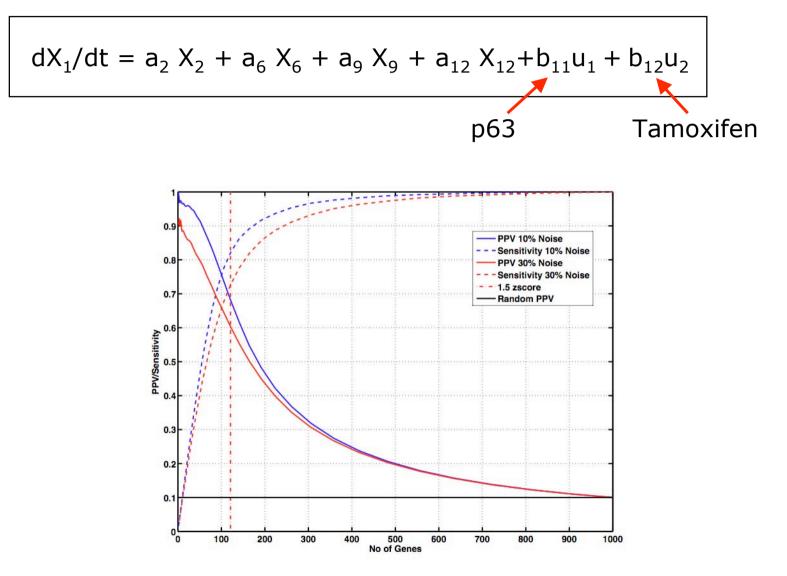
...and Dr Caterina Missero (CEINGE, Napoli) for supervision of experiments; Prof. Tim Gardner and Prof. Jim Collins (Boston University, USA) for MNI/NIR related work.

Mario Lauria



- •First filter
  - Area under curve
- •Second filter
  - Standard deviation is computed at each time point using smoothing and generalized cross validation algorithm
- Statistical test for significance (Chi-Square)







•Integral Approach instead of differential approach

$$\int_{0}^{t_{k}} \dot{X}(t)dt = A \int_{0}^{t_{k}} X(t)dt + B \int_{0}^{t_{k}} U(t)dt \quad k = 1 \dots M$$

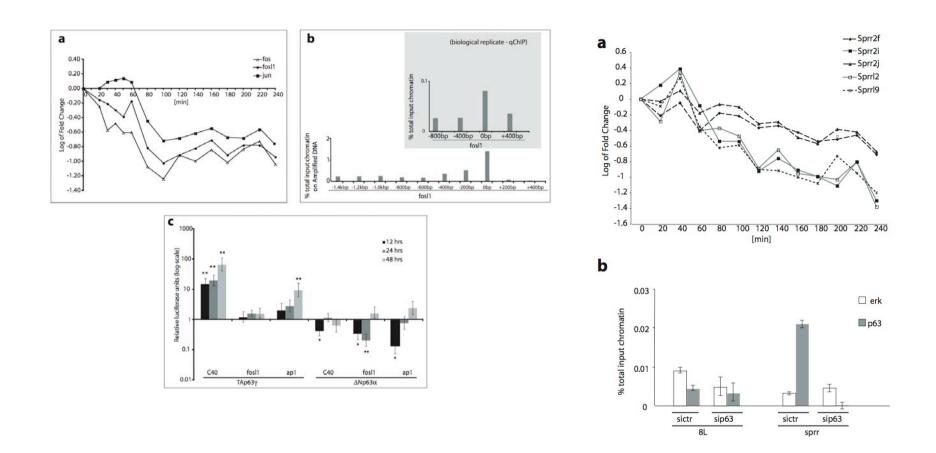
$$X(t_k) = A \int_0^{t_k} X(t) dt + B \int_0^{t_k} U(t) dt \quad k = 1 \dots M$$

•Bootstrapping

Mukesh Bansal, Diego di Bernardo Inference of gene networks from temporal gene expression profiles. IET (Under Review)



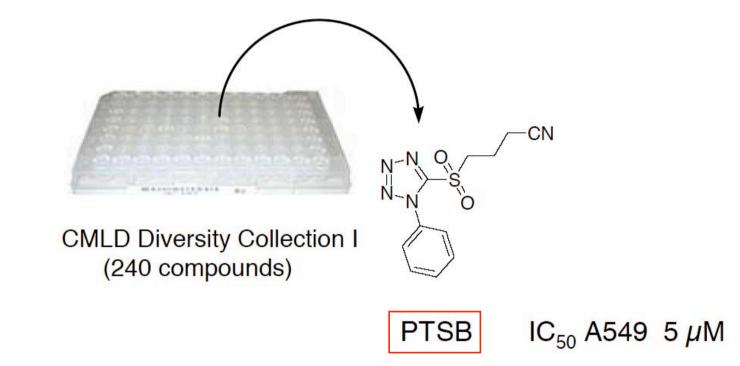
## (1a) Time Series Network Identification Algorithm (NEW):



P63 directly regulates at early times AP1 complex and markers of terminal keratinocyte differentiation *-Manuscript submitted-*



## Identified novel anticancer compound via chemical screen



• PTSB inhibits growth in yeast and tumor cell lines

In collaboration with Schaus and Elliot laboratories Dept. of Chemistry, Boston University Center for Methodology and Library Development (CMLD), Boston U.



# Identifies thioredoxin (TRX2) and thioredoxin reductase (TRR1)

Compound	Known pathway	Known target	Predicted pathway	Ranked target genes (rank)
PTSB	unknown	unknown	cell redox homeostasis	TRRI, TRX2

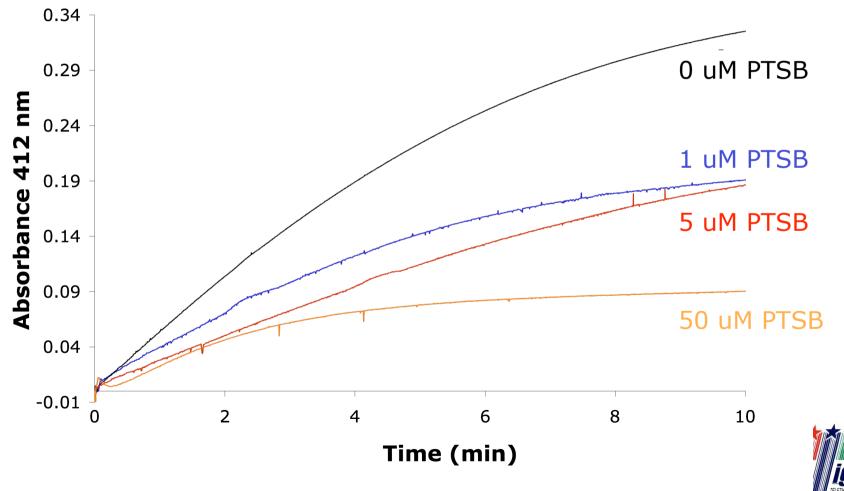


# TRR1/TRX2 activity inhibited in presence of PTSB

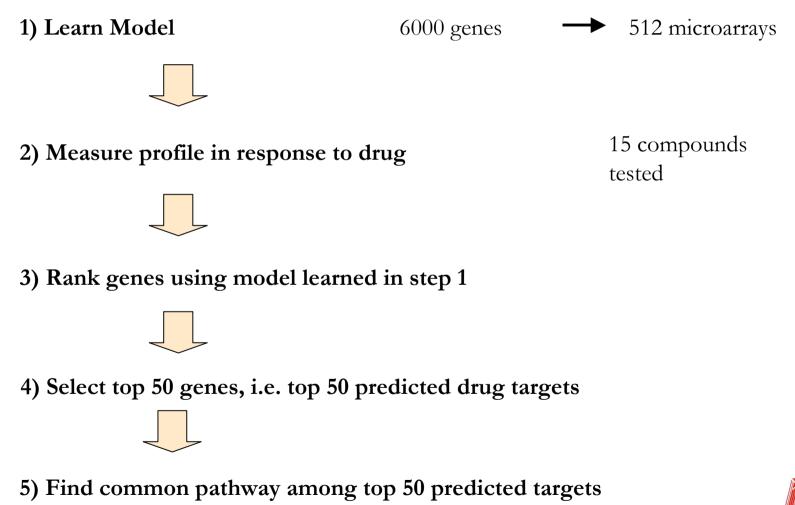
#### Assay:

Thioredoxin reduction of dithio(bis)nitrobenzoic acid (DTNB)

• Product of reaction = thiolate anion, measured via A412

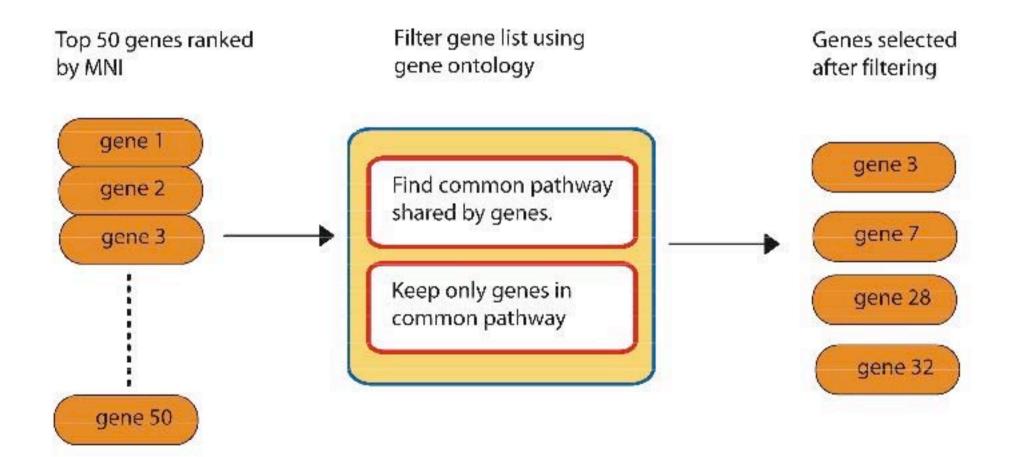


dibernardo.tigem.it

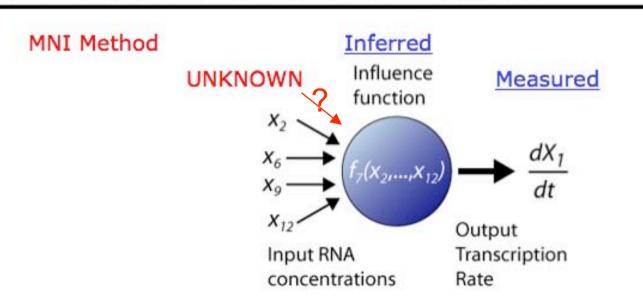




# **Geno Ontology Filtering**

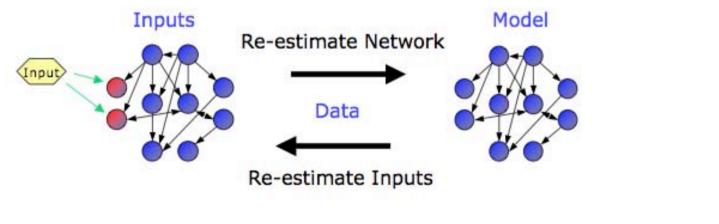


# MNI: Microarray Network Identification



#### The MNI Algorithm: an unsupervised approach

MNI Algorithm: Recursively estimate inputs and model using data

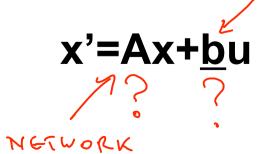


1

$$\begin{cases} \mathbf{x'}_{1}(\mathbf{t}) = a_{11}\mathbf{x}_{1} + a_{12}\mathbf{x}_{2} + \dots + a_{1n}\mathbf{x}_{n} + \mathbf{b}_{1}\mathbf{u} \\ \dots & \text{Drug effect unknown} \\ \mathbf{x'}_{n}(\mathbf{t}) = a_{n1}\mathbf{x}_{1} + a_{n2}\mathbf{x}_{2} + \dots + a_{nn}\mathbf{x}_{n} + \mathbf{b}_{n}\mathbf{u} \end{cases}$$

Or in matrix format:

DRUGEFFECT





 $\mathbf{0} = a_{11}\mathbf{x}_{11} + a_{12}\mathbf{x}_{21} + \dots + a_{1n}\mathbf{x}_{n1} + \mathbf{b}_{1n}\mathbf{u}$ 

 $\mathbf{0} = a_{11}\mathbf{x}_{1h} + a_{12}\mathbf{x}_{2h} + \dots + a_{1n}\mathbf{x}_{nh} + \mathbf{b}_{1}\mathbf{u}$ 

Choose only the *h* experiments where gene 1 has not been perturbed and solve the eqs. for gene 1 with  $a_{11} \neq 0$ .

Repeat for all the genes and obtain matrix **A** 

...we still did not say how to find the *h* experiments where gene *i* has not been perturbed, this is done simply by choosing those experiments where the gene has changed less...



Say  $x_d$  the expression profile following the drug treatment, then the predicted targets of the drug are:

# bu=-Ax<sub>d</sub>



#### Other projects going on in our lab:

- REVERSE ENGINEERING BASED ON MI AND BAYESIAN APPROACH
- IDENTIFICATION OF DRUG TARGETS
- SYNTHETIC BIOLOGY COBIOS (coordinator) 120kE/yr for 3 yrs for my lab

